spenito is required for sex determination in Drosophila melanogaster

Dong Yan* and Norbert Perrimon*,†,1

*Department of Genetics, Harvard Medical School, Boston, MA 02115; and †Howard Hughes Medical Institute, Harvard Medical School, Boston, MA 02115

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Sex-lethal (Sxl) encodes the master regulator of the sex determination pathway in Drosophila and acts by controlling sex identity in both soma and germ line. In females Sxl maintains its own expression by controlling the alternative splicing of its own mRNA. Here, we identify a novel sex determination gene, spenito (nito) that encodes a SPEN family protein. Loss of nito activity results in stem cell tumors in the female germ line as well as female-to-male somatic transformations. We show that Nito is a ubiquitous nuclear protein that controls the alternative splicing of the Sxl mRNA by interacting with Sxl protein and pre-mRNA, suggesting that it is directly involved in Sxl auto-regulation. Given that SPEN family proteins are frequently mutated in cancers, our results suggest that these factors might be implicated in tumorigenesis through splicing regulation.

Sex determination in Drosophila is under the control of the master regulatory gene Sex-lethal (Sxl) (1). Sxl acts downstream of the X-chromosome counting mechanism and encodes a female-specific RNA binding protein. Once activated, Sxl maintains its own expression by regulating the alternative splicing of its pre-mRNA. Sxl controls female fate by controlling somatic and germ-line sex identity as well as dosage compensation (2). In female somatic cells, Sxl controls the alternative splicing of transformer (tra), which together with transformer2 (tra2) controls the alternative splicing of doublesex (dsx) and fruitless (fru). dsx and fru in turn encode sex-specific transcription factors that control male versus female morphology, physiology, and behavior (3, 4). In addition, Sxl represses the male-specific dosage compensation system by regulating male-specific lethal 2 (mstl-2) both at the level of alternative splicing and translational control (5).

In the female germ-line Sxl regulates sex identity by a different mechanism, as tra, tra2, mstl-2 have no roles in the germ line (2). In the ovary, germ-line stem cells (GSCs) located at the anterior tip of the germarium divide to produce another GSC and a cystoblast (CB) that is committed to differentiate. Sxl protein accumulates to high levels in the GSCs/CBs and is required for the proper differentiation of the germ cells (6). Germ cells lacking Sxl cannot differentiate and instead produce stem cell tumors. The identity of Sxl target genes in the germ line is not well characterized; however, a recent study indicates that nanos, a gene required for GSC maintenance, is a Sxl target (7). Indeed, Sxl has been proposed to promote the differentiation of GSCs by downregulating Nanos levels in CBs by binding to the nanos 3′ UTR (7). In addition, Sxl is also important for repressing the expression of testis-specific genes, including Phf7, a male germ-line identity gene (8). In the absence of Sxl, Phf7 is mis-expressed leading to germ-line tumors (9).

Sxl does not act alone to control splicing. Several genes, including sans fille (snf), virilizer (vir), female-lethal-2-d (fl(2)d), SPP45, U1-70K, U2af38, U2af50, and protein partner of sans-fille (pps) facilitate Sxl splicing autoregulation (10–18). Except for pps, these genes encode either general splicing factors or proteins associated with spliceosomes. They all act to maintain the Sxl autoregulatory splicing loop by interacting with Sxl itself. In addition, some of them are involved in the splicing of other Sxl splicing targets such as tra or mstl-2 (1). Interestingly, these genes have essential functions besides Sxl regulation and null mutations are associated with zygotic lethality in both sexes. Therefore, the roles of these factors in sex determination were revealed from genetic interactions (snf, U1-70K, U2af38) (11, 17), temperature-sensitive mutation (vir) (15), clonal analysis (fl(2)d) (10), or biochemical studies (SPP45, U2af50 and pps) (12, 14, 17).

Here, we characterize spenito (nito), a novel regulator of Sxl, which is required to maintain sex identity and Sxl levels in both the female germ-line and somatic tissues. Nito is required for the proper alternative splicing of the Sxl pre-mRNA in both germ line and soma, and forms a complex with Sxl protein and its pre-mRNA, thus identifying an important component of the sex determination pathway.

Results

nito Is an Essential Gene Required for Ovarian GSC Differentiation.

nito was identified from our previous RNAi screen in Drosophila GSCs (19). Specifically, RNAi knockdown of nito driven by the germ-line-specific MTD-Gal4 driver resulted in complete sterility in females. In wild-type ovarioles, two or three GSCs are located in the anterior tip of the germarium (Fig. 1 A–B′). Strikingly, nito shRNA ovarioles are filled with undifferentiated stem-cell-like cells, and nurse cells and oocytes are not formed (Fig. 1 C and C′, compared with WT in Fig. 1 B and B′). Further, stem-cell-like cells associated with nito shRNA ovarioles retain their proliferative potential as shown by staining with the mitotic marker phospho-histone H3 (pH3) (Fig. 1 E, compared with WT in Fig. 1 D). Note that the same stem-cell tumor phenotype was observed with three independent nito shRNAs and two long dsRNA RNAi lines (Methods) (Fig. S1 A–B′), indicating that nito is an essential gene required for GSC differentiation.

nito Is Required for Sex Determination in the Soma.

Because the germ-line phenotype of nito could reflect perturbations in a number of developmental processes affecting either germ-line

Significance

Sex determination is a fundamental biological problem faced by all metazoans. To understand the sex determination pathway, it is important to identify all the genes involved in this process. In this study, we have identified a novel gene, spenito (nito), which is required for sex determination in Drosophila melanogaster. Loss of nito function in the soma transforms female tissues to male, and loss of nito function in female germ-line stem cells changes their sexual identity and prevents them from proper differentiation. We show that nito is a cofactor for Sex-lethal (Sxl) auto regulation, a process that remains an important textbook model for regulated alternative splicing.

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1To whom correspondence should be addressed. Email: perrimon@receptor.med.harvard.edu.

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proliferation or differentiation, we examined nito loss-of-function phenotypes in somatic tissues. Strikingly, expression of nito shRNA using dome-Gal4, that drives expression in both the leg and genital discs (Fig. 2 A and B), led to the transformation of female tissues into that of males. This is evidenced by the appearance of dark, thickened bristles, the male sex combs, in the forelegs of nito shRNA females (Fig. 2E, compare with WT in Fig. 2 C and D). This phenotype is almost fully penetrant and occurs in 97% (n = 78) of females examined. In addition, there are strong abnormalities in the genitalia of these female flies. First, a rotation defect has occurred in 71% (n = 78) of dome-Gal4/nito-shRNA females (Fig. 2H). Second, typical female external structures, such as vaginal bristles (Fig. 2G, white arrow), are absent in the genitalia (Fig. 2H). Third, structures resembling those of males, such as penis apparatus and claspers can be identified (Fig. 2 F and H). These transformations suggest that Nito is a component of the Drosophila sex determination pathway in the soma. Because Sxl shRNA generates a stem-cell-tumor phenotype in the germ line similar to that of nito (Fig. 1 F and F'), the nito germ-line phenotype therefore could be due to sex determination defects associated with Sxl (see below).

Nito Is a Ubiquitously Expressed Nuclear Protein That Is Crucial in Both Sexes. Nito, together with Split ends (Spen), are members of the SPEN protein family characterized by three N-terminal RNA recognition motifs (RRMs) and a C-terminal SPOC (Spen paralog and ortholog C-terminal) domain (Fig. 3A) (20, 21). To analyze Nito expression, we raised a polyclonal antibody against a 22 amino acid peptide (Methods). A Western blot showed that this antibody recognizes a protein of the expected ~89 kDa size in Drosophila S2 cell lysates (Fig. 3C). Nito is ubiquitously expressed in all tissues examined, including imaginal discs and ovaries, and localizes to the nucleus (Fig. 3 D and F). Furthermore, expression of nito shRNA using ap-Gal4 led to almost complete depletion of the Nito protein in the dorsal half of wing discs demonstrating the specificity of the antibody (Fig. 4C and Fig. S2 A and B).

Because nito affects sex determination, we tested whether its expression level is biased in females versus males. To exclude the maternal contribution from ovaries, we compared nito mRNA levels in wing discs. As shown in Fig. 3B, nito mRNA levels were similar in female and male wing discs. In addition, Nito antibody staining showed similar protein levels in female and male discs (Fig. 3 D and E). Further, Nito is not regulated by Sxl as its protein level is not affected in Sxl RNAi discs (Fig. 3 G and G'). Together, these data indicate that nito is not differentially expressed in males versus females.

We generated a null allele of nito by imprecise P-element excision (referred to as nito1) to test whether nito is an essential gene. nito1 homozygous animals die during larval stages and homozygous mutant clones show the absence of Nito protein indicating that nito1 is a null mutation (Fig. 3 H and H'). Interestingly, nito1 causes lethality in both females and males, indicating that nito is an essential gene. Further, nito1 is lethal over

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**Fig. 1.** Nito is essential for ovarian GSC differentiation. (A) Diagram showing the structure of a wild type germarium. (B–B') WT ovarioles stained for α-Spectrin, Vasa and DAPI. The α-Spectrin antibody labels the round spectrosomes in GSCs (arrow); the Vasa antibody labels all germ cells and DAPI labels nuclei to monitor proliferation or differentiation, we examined nito loss-of-function phenotypes in somatic tissues. Strikingly, expression of nito shRNA by MTD-Gal4 were stained for α-Spectrin, Vasa and DAPI. Note the numerous stem-cell-like cells labeled by α-Spectrin and the absence of differentiated nurse cells. (D–E) pH3 staining in WT egg chambers and egg chambers expressing nito shRNA. In WT egg chambers, pH3-positive cells were restricted to the anterior tip of the gerarium but were detected throughout nito shRNA egg chambers (arrowheads). (F–F') Egg chambers expressing Sxl shRNA stained for α-Spectrin, Vasa and DAPI. (Scale bars: 20 μm.)
Nito interacts with Sxl protein and its pre-mRNA in S2 cells. We then analyzed how Nito controls Sxl alternative splicing. Because the key protein that binds Sxl pre-mRNA and inhibits splicing of male-specific exon 3 is Sxl itself (2), we examined whether Nito interacts with Sxl using a coimmunoprecipitation (co-IP) assay in Drosophila S2 cells. GFP-Nito and HA-Sxl were expressed either individually or in combination in S2 cells, and GFP alone was used as a control (Fig. 5A). HA-Sxl was detected in the precipitate obtained using anti-GFP nanobody from GFP-Nito cells, but not from GFP-expressing cells. Similarly, GFP-Nito, but not GFP, was pulled down by HA-Sxl. Further, we tested whether the interaction between Nito and Sxl is dependent on the presence of RNA. Interestingly, the amount of Sxl-HA pulled down by GFP-Nito is strongly reduced in the presence of RNase, suggesting that the Nito/Sxl interaction is mediated or stabilized by RNA (Fig. 5B). Finally, we performed RNA immunoprecipitation (RIP) experiments in S2 cells to analyze whether Nito can interact with Sxl pre-mRNA, which was detected by RT-PCR using an intron 3-exon 4 primer pair (12). As shown in Fig. 5C, GFP-Sxd, GFP-Nito, but not GFP alone, can pull down Sxl pre-mRNA from cell lysates. Together, the specific interactions between Nito, Sxl, and Sxl pre-mRNA support the model that Nito forms a complex with Sxl and that they together regulate alternative splicing of Sxl mRNA (Fig. 5D).

Screen for Additional Splicing Genes Involved in Sex Determination. Because we identified nito as an important gene involved in sex determination by regulating Sxl splicing, we asked whether there are other unidentified genes in the Drosophila genome acting in this pathway. Because most known Sxl-autoregulatory proteins are associated with spliceosomes (2), we screened a collection of 316 RNAi lines representing 247 splicing-associated genes and RNA-binding proteins (Dataset S1), using MTD-Gal4 for the germ line and dome-Gal4 for somatic phenotypes. In addition, we also included in our screen a wing-specific driver, nub-Gal4, as many RNAi lines exhibit lethality with dome-Gal4 and prevent the detection of potential sex determination phenotypes. Because Sxl is responsible for the larger wing size in females, examination of wing size using nub-Gal4 allows detection of potential genes involved in sex determination, as shown in the case of nito (Fig. S3 A–F).

Our screen successfully identified known components of the sex determination pathway: tra and tra2 RNAi showed strong female-to-male transformation in sex combs and genitalia when induced by dome-Gal4; and f(2)2d and vir were identified using the nub-Gal4 driver as both have a stronger effect in female wings than in male wings. Strikingly, besides these genes we did not identify any other genes that showed a sex-related phenotype. However, we did characterize GSC differentiation phenotypes associated with tsu, mago, RupS1, Rbp9 RNAi lines, as well as a deficiency of the nito locus indicating that the lethality is likely due to the nito mutation. Consistent with this, nito RNAi driven by a ubiquitous Gal4 such as actin-Gal4 or tubulin-Gal4 is associated with larval lethality.
as wing growth and pattern defects with *kul, CG7879, ASPP, tst* and *Syp* RNAi lines. These data provide a valuable resource of phenotypes associated with splicing-related genes (data available at www.flyrnai.org/RSVP.html).

**Discussion**

We describe the characterization of Nito as a novel component of the *Drosophila* sex determination pathway. Nito loss-of-function results in stem-cell tumor phenotypes in the germ-line and sexual transformations in the soma. Interestingly, Nito affects Sxl protein levels in both GSCs and somatic tissues by regulating Sxl pre-mRNA alternative splicing, most likely directly as Nito interacts with the Sxl protein and pre-mRNA. The role of Nito is reminiscent of the previously reported roles of splicing factors in Sxl auto regulation, such as both subunits of U2AF (17), U1-70K (17, 18), Fl(2)d (10, 23), SPF45 (13, 14), Vir (15, 24), and Snf (11, 16). Our data support earlier reports that Sxl physically interacts with components of the spliceosome to simultaneously block utilization of the 3′ and 5′ splice sites of the male exon.

Nito and Spen are members of the SPEN protein family that are evolutionarily conserved from plants, worms, flies to mice and humans (25). Both proteins contain three N-terminal RRM domains and one C-terminal SPOC domain. The sequence similarity between these domains is low and there is no conservation outside these motifs, suggesting that they have evolved specific functions following a duplication event (20), as indicated by our observation that *spen* is not required for *Sxl* regulation (Fig. S2C). In *Drosophila*, *spen* was first identified in several genetic screens looking for components of the receptor tyrosine kinase (RTK) signaling pathway (26). Subsequent studies found
Genetic studies in *Drosophila* have shown that *nito* overexpression results in a rough eye phenotype (20) and that it plays a redundant role with *spen* in Wnt signaling (21), but how Nito is involved in these processes is not known. Biochemical studies indicate that Nito, like its human ortholog, copurify with the precatalytic splicingosome (complex B) (34). In addition, *nito*, as well as many other splicing factors, was identified in an RNAi screen for RAS/MAPK signaling components (35). Consistent with these findings, we find that *nito* is required for the alternative splicing of the master sex-determination gene *Sxl*. Previously, both Spen and Nito were thought to act mainly as transcription factors through their SPOC domains, our findings however clearly indicate that Nito is involved in mRNA splicing. It is intriguing to note that PPS, another important factor required for *Sxl* splicing, also has a SPOC domain (12). Similar to Nito, PPS also forms a complex with Sxl protein and its pre-mRNA (12). In the future it will be crucial to dissect how different protein domains contribute to the function of SPEN family proteins.

Then what is the “main” role of *nito*? On one hand, the phenotypes in the sex comb, genitalia and germ line appear specific to *Sxl* and such phenotypes do not depend on the genetic interaction with other genes in the sex determination pathway. On the other hand, *nito* clearly has other non-sex-specific functions, as revealed by the lethality, rough eye, and wing phenotype observed in both sexes (Fig. S3 A–F). Because a null allele of *nito* is associated with zygotic lethality, the RNAi knockdown approach is a powerful method to reveal sex-related phenotypes. Interestingly, our RNAi screen targeting splicing factors did not identify any new additional sex determination genes, indicating that there are a limited number of genes yet to be identified in this pathway. Finally, intriguingly, three recent studies have identified SPEN and Rbm15 (the mouse and human ortholog of Nito) as factors interacting with *Xist*, the long noncoding RNA that is essential for dosage compensation in mammals (36–38). Clearly, future experiments such as RNA-seq will be necessary to elucidate the mechanism and logic of Nito-mediated signaling events.

Rbm15, also known as OTT, was originally identified from infants with acute megakaryoblastic leukemia (AMKL) (39, 40). The t(1, 22) chromosomal translocation results in fusion of

![Fig. 4. Nito is required for Sxl levels and regulates Sxl mRNA splicing. (A and B) Sxl stainings in WT male (A) and female (B) wing discs. (C–C′) Expressing nito shRNA in the dorsal half of the disk (below the dashed line) using ap-Gal4 leads to strong reduction of Nito (C) and Sxl (C′) stainings. (D–D′) Sxl antibody staining (D′) in wing discs containing nito1 mutant clones, which are marked by the absence of GFP (D). Note the absence of Nito and Sxl staining in nito1 mutant clones. (E and F) Sxl stainings in WT egg chambers (E) or in egg chambers expressing nito shRNA by MTD-Gal4 (F). Scale bars: 20 μm. (G) Diagram showing the alternative splicing event that produces the male- or female-specific Sxl transcripts. The arrows indicate the primers used for RT-PCR. Sxl splicing was analyzed by RT-PCR using RNA extracted from wing discs or ovaries. Male-specific bands: 2–3-4. Female-specific bands: 2–4.

![Fig. 5. Nito interacts with Sxl and Sxl pre-mRNA in S2 cells. (A) HA-Sxl, GFP-Nito or GFP expression vectors were transfected individually or together into Drosophila S2 cells. Cell lysates were immunoprecipitated using GFP nanobody or anti-HA antibody and analyzed by Western blot. GFP alone is used as a control. Asterisk indicates IgG heavy chain. (B) S2 cells were transfected with GFP-Nito and HA-Sxl, and Co-IP was performed using GFP nanobody in the absence or presence of RNase A and RNase T1. (C) S2 cells were transfected with GFP-Nito, GFP-Sxl or GFP and immunoprecipitated with GFP nanobody. The presence of Sxl pre-mRNA was detected by RT-PCR using an intron 3/exon 4 primer pair. GFP-Sxl was used as a positive control and GFP alone as a negative control. (D) Model: Nito forms a complex with Sxl and together they repress the splicing of Sxl exon 3 in female tissues.](https://www.pnas.org/doi/10.1073/pnas.1515891112 Yan and Perrimon)
RBMI5 and MKL1, and the fusion protein is responsible for AMKL development as shown in a mouse model (41). In addition to this chromosome translocation, recent cancer genome sequencing projects have found that RBMI5 and SPEN (also known as SHARP) are mutated in many different types of cancers, such as adenoid cystic carcinomas and bladder cancers (42). Given that SPEN family proteins are frequently mutated or deleted in cancers, they have been proposed to act as potential tumor suppressors (42). Studies of Spen and Nito in Drosophila will provide mechanistic insights to our understanding of this important family of proteins.

**Methods**

Details on the fly strains used in this study, as well as how the null nito mutation was isolated and how nito clones were generated can be found in **SI Methods**. Protocols used for antibody staining, reagents, how Nito antibodies were generated, commounprecipitation protocols, RT-PCR, and information on primers and RNA immunoprecipitation (RIP), can be found in **SI Methods**.

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Supporting Information

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SI Methods

Fly Strains. The following stocks were used in this study: w1118 (used as wild-type, WT), MTD-Gal4, ap-Gal4, nob-Gal4, dome-Gal4, UAS-2xEFFP (43). Sxl shRNA (HMS00609), nito shRNA (HMS00166), nito shRNA (HMS02013), nito shRNA (HMJ02081), nito dsRNA (VDRC 20942), nito dsRNA (VDRC 114704). Experiments presented in Figs. 1, 2, and 4 and Fig. S3 were done using nito shRNA (HMS00166).

To generate a null nito mutation, we used the homologous viable P-element insertion nitoHP25329 located in the 5’UTR of the nito gene. After mobilization of the HP25329 P-element, we screened for homologous lethal lines and recovered a null allele, nito1, that deletes 1,357 bp and is lethal over Df(2R)Exel6055 (43F1 to 44A4) that uncovers the nito locus. The Nito antibody, raised against amino acids 479–500, cannot detect any Nito antigens in nito1 mutant clones (Fig. 3 H and H’).

To generate mutant clones, nito1 was recombined with FRT413 and crossed to y w hsflp; ubiGFP FRT413 flies. The progeny were heat-shocked at 37 °C for 1 h twice at first- and second-instar larval stage.

Antibody Stainings in Discs and Ovaries. Larval wing discs and female ovaries were stained as described (19). Briefly, tissues were dissected in PBS and fixed in 4% formaldehyde in PBST (PBS + 0.1% Triton X-100). After blocking in 1% normal donkey serum in PBST for 1 h, the samples were incubated with the primary antibody in the same solution at 4 °C overnight. After three washes in PBST, samples were incubated with the secondary antibody for 2 h at room temperature, washed in PBST three times, and subsequently mounted in Vectashield. All images were taken on a Zeiss LSM 780 microscope.

The following antibodies were used: mouse anti-α-Spectrin (1:10) (3A9, DSHB), rabbit anti-Vasa (1:250) (Santa Cruz Biotechnology), mouse anti-Sxl (1:10) (M18, DSHB), rabbit anti-Nito (1:500), rabbit anti-phospho-Histone H3 (1:1,000) (Millipore), rabbit anti-GFP (1:1,000) (Molecular Probes), mouse anti-GFP (1:200) (Molecular Probes), Alexa 488- or 555-conjugated secondary antibodies (1:1,000) (Molecular Probes) and DAPI (1:1,000) (Molecular Probes).

Nito antibodies were generated in rabbits against a peptide containing amino acids 479–500 (KSSKPyDESALEYRRPEYDPY) and affinity-purified at YeniZymes Antibodies. Polyclonal antisera were raised in two rabbits, YZ3137 and YZ3138, and gave similar staining patterns. All of the experiments described in the paper were performed with anti-Nito.

Adult legs and wings were mounted in a 1:1 (vol/vol) mixture of Permount (Fisher Scientific) and xylene. The genitalia images were taken in stacks and rendered with HeliconFocus software.

Coimmunoprecipitation. To generate the GFP-Nito plasmid, a nito full-length cDNA (GH11110) was cloned into the Drosophila Gateway vector pAGW. HA-Sxl and GFP-Sxl were constructed following PCR of Sxl (the MS3 isoform) from UAS-Sxl flies (44) and cloned into pAHW and pAGW, respectively. GFP was cloned into pAWM as a control.

Drosophila S2 cells were maintained at 25 °C in Schneider’s medium supplemented with 10% FBS. One microgram of total DNA was transfected into S2 cells in a single well of six-well plates with Effectene (Qiagen). After 48 h, cells were lysed in IP lysis buffer (Pierce) with Halt Protease Inhibitor (Thermo Scientific). Lysates were incubated with anti-GFP nanobody agarose beads (Allele Biotechnology) or anti-HA agarose (Sigma) for 2 h at 4 °C. The beads were washed 3–4 times with 1 mL lysis buffer. Protein complexes were eluted and detected by Western blotting using anti-GFP antibody (A6455, Molecular Probes) or anti-HA antibody (3F10, Thermo Scientific) were added to 1 mL of lysis and incubated for 30 min at 30 °C, then overnight at 4 °C with beads (12, 17).

RT-PCR. Total RNA was extracted from dissected wing discs or ovaries using TRIzol (Invitrogen), digested with DNase I (Qiagen) and purified using the RNAeasy Mini kit (Qiagen). cDNA was generated from 1 μg of purified RNA using the iScript cDNA Synthesis kit (Bio-Rad). For nonreal time PCR, TaKaRa Taq polymerase was used. For qPCR, iQ SYBR Green Supermix (Bio-Rad) was used and reactions were measured in a CFX96 Real-Time PCR detection system (Bio-Rad). qPCR results for nito expression in male and female wing discs (Fig. 3B) were normalized to the reference gene αTubulin84B. Sxl primers used in Fig. 4G are described in ref. 12. nito primers in Fig. 3B and αTubulin84B primers are listed below.

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<th>αTubulin84B-f</th>
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RNA Immunoprecipitation (RIP). RIP experiments were performed following previous protocols (45, 46). One microgram of DNA was transfected into S2 cells in 60-mm plates with Effectene (Qiagen). After 48 h, cells were lysed in IP lysis buffer (Pierce) with Halt Protease Inhibitor (Thermo Scientific) and RNasin plus (40 U/mL, Promega). After centrifugation, 10% of the supernatant were saved as the input and the rest were incubated with anti-GFP nanobody agarose beads (Allele Biotechnology) for 2 h at 4 °C. The beads were washed 3–4 times with 1 mL of lysis buffer. To elute RNA from the protein complexes, the beads were treated with protease K solution for 10 min at 55 °C. Total RNA from the input and the beads were extracted by using RNaseasy Micro kit (Qiagen). cDNA was synthesized using SuperScript III First-Strand Synthesis System (Life Technologies) with four of the eluted RNA and random hexamers. TaKaRa Taq polymerase was used for two rounds of PCR amplification with primers and conditions described in ref. 12.

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<th>Sxl-intron</th>
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Fig. S1. Two independent nito shRNAs result in similar stem-cell-tumor in the germ-line. (A–B’) Egg chambers expressing shRNAs targeting nito (HMJ02081) or nito (HMS02013) using MTD-Gal4 stained for α-Spectrin, Vasa and DAPI. (Scale bars: 20 μm.)

Fig. S2. nito, but not spen, regulates Sxl levels in wing discs. (A–B’) Expression of nito shRNA (HMJ02081 or HMS02013) in the dorsal half of the wing disk using ap-Gal4 leads to a strong reduction of both Nito (A and B) and Sxl (A’ and B’) stainings. (C) Expression of spen shRNA in the dorsal half of the disk (below the dashed line) by ap-Gal4 does not affect Sxl protein levels. spen shRNA generates embryonic lethality with cuticle and head defects when expressed using MTD-Gal4 (47), which resembles the phenotype of the spen mutant, indicating that the shRNA is functional.

Fig. S3. Nito shRNA affects wing growth more strongly in females than in males. (A) WT male wing. (B) WT female wing. (C) Overlay of the images in A (red) and B (blue) shows that a WT female wing is about 30% larger than a WT male wing. Male (D) and female (E) wings in which nito shRNA was expressed using the nub-Gal4 driver. (F) Overlay of D and E showing that both male and female wings reach about the same size upon nito knockdown.

Dataset S1. Screen results of 316 RNAi lines targeting 247 splicing genes using MTD-Gal4, dome-Gal4 and nub-Gal4